# Total Phenolic, Cinnamic Acids and Selected Microelements in Gluten Free Pasta Fortified with Banana

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The aim of this study is to obtain gluten free pasta fortified with 10-40% fresh banana and dried banana in absence and in presence of ascorbic acid. Also, the obtained products were studied in term of total phenolic, cinnamic acids and selected microelements (Fe, Cu, Zn, Mn, Ni) composition. The results showed that addition of 30-40% fresh or dried banana in gluten free pasta leads to increasing of bioactive compounds of products.

Keywords: gluten free pasta, polyphenols, cinnamic acids

Gluten-free products are intended for people with celiac disease and who have a permanent intolerance to gluten and are obtained from grain of which proteins are not gluten generators. Rice is a grain matrix used successfully both in celiac diet and in technology of obtaining pasta. Rice pasta however, presents a low macro and micronutrients necessary for the body development. Gluten free pasta contents less phosphorus, potassium, calcium, magnesium, iron and zinc as regular pasta made from wheat.

In obtaining of a functional and dietary product an important role plays the biologically active compounds in its composition. Grains are an important source of elements and chemical compounds with potential biological activity, such as phenolic compounds [16, 18]. Phenolics in cereals can be subdivided into acids derived from either benzoic acid or cinnamic acids (CA). The most commonly CA found in cereals have seven carbon atoms, a benzenic ring, a carboxylic group and one or more hydroxyl and/or methoxyl groups [10]. Ferulic, p-coumaric, caffeic and sinapic acids are the main CA [10,18]. Ferulic acid represents one of the most abundant CA in grains [3,6,26]. CA presents biological importance having antioxidant anti-inflammatory and anticancer properties. Also is involved in reducing the level cholesterol and triglycerides and implicitly the risk of cardiovascular disease [16].

The aim of fortification is to increase the nutritional value of product by supplying with nutrients or biological active compounds. The fortification of pasta with fruits is relatively unusual. Combining the effect of rice gluten-free pasta with banana is an opportunity to obtain a novel dietary food with nutraceutical high potential [14,20,21].

Nutraceutical properties of bananas have been reported in previous studies [5,7,22]. The high content in fibre, polyphenols and potassium recommends the bananas consumption in food diet [9,19]. The use of ascorbic acid as pre-treatment in pasta producing technology increase the biological value of product and prevents enzymatic browning, biochemical and microbial activities that occur during pasta production [4].

The purpose of this study is: i) to obtain three types of gluten-free pasta made of rice flour (RF) with addition of

different proportions of dried fruits of banana (DB), fresh banana (FB) and fresh banana pre-treated with ascorbic acid (FBA), ii) to characterize the products obtained, in terms of polyphenols, cinnamic acids, macro and micronutrients content.

## **Experimental part**

Materials and methods

Raw materials

Rice flour (RF) was obtained from Pirifan, Romania, dried banana from Poex Czech and fresh banana from Kaufland market. In case of obtaining pasta with ascorbic acid addition, acid was added previously to fresh bananas, which were maintained 30 minutes in acid medium before introducing in the manufacturing recipe.

## Pasta formulation

Pasta was prepared with rice flour by adding different concentrations (10, 20, 30 and 40% w/w) of DB, FB and FBA. One egg and 10 mL water were added for each sample. The dough was manually formed and cut in a pasta machine (Pasta machine, Kitchen collection, WN16195-150, China). Obtained pasta were dried 24 h at room temperature.

#### Microelements determination

The determination of microelements was done by the flame atomic absorption spectroscopy method (AAS) with graphite furnace. 3 g samples were burned 6 h at 650°C in furnace (Nabertherm B150, Lilienthal, Germany). The ash was dissolved in HCl 20%. Standard solutions (ICP Multielement Standard solution IV CertiPUR) were purchased from Merck. Each value is the mean of three independent determinations.

Total phenolic content (TP) and cinnamic acids (CA) determination

Total phenolic content was determined using a modification method of Arranz, 2010 [6]. Briefly, 0.5 g pasta was extracted 1 h at room temperature with 20 mL methanol:water (50:50, v/v) and the pH was adjusted to 2 with acetic acid.. The samples were centrifuged 10 min at 450 rot/min. 0.5 mL supernatant was treated with 1.25 mL

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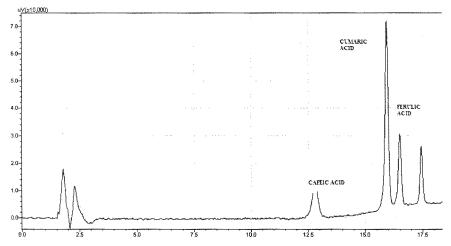


Fig. 1. HPLC chromatogram of cinammic acids (cafeic, ferulic, cumaric).

Cua Nie Fe Zna Samples Mn 1.984±0.006 15.801±0.023 16.030±0.023  $0.556 \pm 0.042$  $5.641\pm0.042$ DB 1.623±0.005 0.529±0.015 12.314±0.021  $20.539 \pm 0.012$  $11.032 \pm 0.041$ 10 % DB  $0.848 \pm 0.037$ 0.434±0.013 3.852±0.027 8.426±0.015 8.448±0.062 20 % DB 0.405±0.006  $1.506\pm0.013$  $5.734\pm0.015$  $17.153\pm0.046$ 15.056±0.055 30 % DB 2.172±0.014 0.345±0.030 8.313±0.022 23.930±0.022 18.293±0.175 40 % DB 2.663±0.018 0.355±0.046 10.640±0.022 18.045±0.023 15.560±0.386 10 % FB 1.906±0.152  $0.383 \pm 0.022$ 5.639±0.063 14.562±0.289 14.887±0.132 20 % FB 1.652±0.135  $0.253 \pm 0.116$ 6.205±0.225 14.414±0.208 16.286±0.112 30 % FB 2.053±0.092 0.432±0.041 6.888±0.125 13.096±0.140 15.770±0.284 40 % FB 1.678±0.024 0.456±0.037  $6.443 \pm 0.323$ 10.170±0.116 15.346±0.186 10 % FBA 1.508±0.022 0.268±0.024 5.239±0.222 10.003±0.057 12.673±0.308 20 % FBA 1.455±0.023  $0.109 \pm 0.052$ 2.613±0.256 10.159±0.153 11.611±0.175 30 % FBA  $1.818\pm0.024$  $0.129\pm0.022$ 4.181±0.171 9.926±0.018 11.450±0.150 40 % FBA  $1.888 \pm 0.055$ 0.131±0.018 3.896±0.497 13.735±0.192 11.486±0.104

Table 1
MICROELEMENTS
CONTENT (PPM) IN
FORTIFIED GLUTEN FREE
PASTA

Folin-Ciocalteu reagent diluted 1:10 with water. The sample was incubated 5 min at room temperature and 1 mL Na $_2$ CO $_3$  60g/Lwas added. After 30 min of incubation at 50 $^{\circ}$ C the absorbance of samples were measured at 760 nm. Results were expressed as  $\mu g$  gallic acid equivalents per g sample ( $\mu g$  GAE/g). Linearity was obtained between 5–50  $\mu g$ /mL.

The main CA in pasta (ferulic, caffeic and cumaric acids) was determined using high performance liquid chromatographic method (HPLC). LC-Shimadzu chromatograph equipped with SPD-10A UV detector, PERVAIL column 150 mm, internal diameter 4.6 mm was used. The chromatographic conditions were: mobile phases A: CH<sub>2</sub>OH:CH<sub>2</sub>COOH:H<sub>2</sub>O (90:2:8, v/v), B: CH<sub>2</sub>OH:CH<sub>3</sub>COOH:H<sub>2</sub>O (10:2:88, v/v), gradient program: 0 min. B 100%, 10 min B 85%, 15 min B 50%, 20 min B 30%, 25 min B 80%. The chromatogram of CA standard is presented in figure 1.

## Results and discussions

Microelements content

In table 1 are presented the results obtained regarding microelements composition (Mn, Cu, Ni, Zn, Fe) of pasta with DB, FB and FBA.

Microelements have a fundamental role in the human diet. Celiac disease is often associated with mal absorption of the nutrients, vitamins and minerals in the gastro intestinal tract. In gluten free products the content of minerals varied considerably among the types of foods. Previous studies reported in gluten free pasta iron content 26 ppm, zinc 8 ppm and copper 4 ppm [25].

In our study the content of microelements in fortified pasta follows the sequence Fe >Zn>Mn>Cu>Ni.

Iron is important for the synthesis of hemoglobin and myoglobin and for the formation of iron-containing enzymes

involved in electron transfer and oxidation-reductions [1]. Iron deficiency represents the most widespread and important micronutrient deficiency in humans [17]. Iron content in fortified pasta varies between 8.426-23.930 ppm, the highest content was detected in pasta with 30%DB. Pasta obtained with FBA in different proportions contain iron in lower amounts than pasta obtained with the FB addition without acid treatment, probably due to the reduction of ferric iron to ferrous iron and binding in soluble complexes [1]. Inclusion of ascorbic acid in recipe of gluten free pasta represents an efficient technological method to enhance the iron absorption, literature studies highlighting the role of acid in bioavailability of iron [1, 13].

Copper content of pasta varies between 0.848 to 2.663 ppm being lower compared with data reported in the literature for gluten-free pasta [25]. Maximum copper content (2.663 ppm) was registered in the sample with 40% DB addition. Higher copper content than control sample was registered also in samples with addition of 30% DB and 30% FB.

Manganese is a component of metal enzyme complexes and acts as an activator in reactions catalyzed by enzymes [12]. Manganese content of bananas (12.314 ppm) is higher than of rice flour (5.641 ppm). Fortification with 40% DB leads to double increase of the manganese content (10.64 ppm) compared with the control sample.

Nickel enters into the human body through primary food and drinking water. In terms of Ni content, our results indicate no significant differences between the values obtained for the control sample and sample with DB. Fortification pasta with fresh or dried banana does not influence nickel intake in the diet. Nickel content is between 0.109-0.556 ppm and is the microelement analyzed that is found in the lowest concentrations in banana pasta studied.

ameans  $\pm$  standard deviation (n=3).

Samples	$TP^a$	Caffeic Acid <sup>a</sup>	Cumaric acida	Ferulic acida
	μg/g	μg/g	μg/g	μg/g
RF	242.10±0.42	8.68±0.01	30.47±0.06	43.8±2.43
DB	$244.00\pm1.41$	2.15±0.03	$10.85\pm0.04$	14.79±0.43
10 % DB	196.95±0.07	$6.79\pm0.01$	$7.28\pm0.05$	13.66±1.53
20 % DB	204.45±0.35	$7.52\pm0.01$	13.28±0.33	22.34±0.57
30 % DB	$213.80\pm0.56$	6.11±0.06	$17.20\pm0.12$	30.41±0.52
40 % DB	$227.80\pm0.40$	6.19±0.01	12.57±0.33	28.44±0.48
10 % FB	$306.86 \pm 0.11$	14.33±0.35	21.76±0.87	24.83±0.85
20 % FB	329.23±0.25	10.03±0.35	25.40±0.62	37.43±0.66
30 % FB	$365.23\pm0.25$	8.46±0.40	$14.80\pm0.80$	23.56±0.81
40 % FB	$367.40\pm0.35$	$10.30\pm1.4$	10.76±0.45	31.40±0.34
10 % FBA	$328.06\pm0.11$	5.36±0.49	10.00±0.35	50.13±1.00
20 % FBA	408.83±0.06	14.70±0.65	8.70±0.36	52.73±0.64
30 % FBA	490.46±0.11	12.06±0.30	10.03±0.35	27.20±0.20
40 % FBA	509.13±0.11	$9.86 \pm 0.41$	13.30±0.36	22.63±0.47

Table 2
TOTAL PHENOLICS (TP) and
CINNAMIC ACIDS (CA) CONTENT
(μg/g) IN FORTIFIED GLUTEN
FREE PASTA

ameans  $\pm$  standard deviation (n=3).

Zinc represents an essential metal entering into the composition of numerous metal enzymes. Whole cereals are good sources of zinc (20-78 ppm) [15], while in glutenfree products zinc content reported in the literature is lower (8 ppm) [25]. The zinc content in the samples analyzed varies between 8.448-18.293 ppm, the maximum values were registered in the case of the 30% DB addition. Fresh or dried bananas fortification leads to increased zinc intake of pasta, while acid treatment applied to fresh bananas induces a decrease in zinc content compared with the control sample.

#### TP and CA content

In table 2 are presented the results obtained regarding TP and CA (ferulic, cumaric and caffeic) of pasta with DB, FB and FBA.

TP content varies between 196.95-509.13 µg/g, the maximum values recorded being registered in the case of the ascorbic acid addition in the pretreatment of fresh bananas (328.06-509.13 µg/g), and the minimum values in the case of pasta with dried bananas (196.95-227.8 µg/ g). TP content of pasta obtained from rice flour, determined in this study, was in agreement with previous published works and highlighting the TP content in various products derived from cereals between 14 and 543 µg/g depending of the nature of the extraction environment and analyzed matrix [2,11]. Verma, 2009 highlights a higher content in TP (1509.0-1306 mg/g) in different wheat varieties [26] and in oat based breakfast cereals, TP content ranged from 1506 to 1853 μgGAE/g [23]. In term of TP our results recorded in pasta fortified with DB are lower than the values recorded in pasta fortified with FB. The addition of banana in pasta induce an increase of TP content, the increase being significant when bananas are pretreated by adding ascorbic acid which also acts as antioxidants. Similar results regarding the increase of the TP in bananas treated with organic acids have been reported by Anyasi [4]. The way in that leads the hydrolysis is particularly important in CA recovering from cereals. Arranz, 2010 highlights the role of acid hydrolysis [6], while Verma, 2009 [26] highlights the importance of alkaline environment. Our results shown the increase of total polyphenol content in the case of ascorbic acid pre-treatment.

Regarding ČA content the level of CA varies in order: ferulic>cumaric>caffeic. Ferulic acid is the most abundant cinnamic acid in cereals, representing up to 90% of total phenolic compounds [8].

CA is predominantly assigned in the coating and less in the grain endosperm. The literature studies reported a content of 11.5  $\mu$ g/g ferulic acid in bran respectively 2.8  $\mu$ g/g in wheat flour [6]. Our results highlight the high ferulic acid content in the control sample (43.8  $\mu$ g/g) and lowest in DB (14.79  $\mu$ g/g). In pasta samples with DB, ferulic acid

varies between 13.66-30.41  $\mu$ g/g, coumaric acid between 7.28-17.20  $\mu$ g/g, and caffeic acid between 6.11-7.52  $\mu$ g/g, the maximum values being recorded in the case of the 30% DB addition for coumaric and ferulic acid and to 20% DB addition of caffeic acid.

FB brings additional intake of CA. So, the caffeic acid content varies between 14.33-8.46  $\mu$ g/g, coumaric acid between 10.76-25.40  $\mu$ g/g, and ferulic acid between 24.83-37.43  $\mu$ g/g. The pretreatment in acid medium of bananas does not lead to a significant change regarding the caffeic and coumaric acid content, instead there is an increase regarding ferulic acid content, which in the case of 20% FB addition increase up to 52.73 $\mu$ g/g.

## **Conclusions**

Banana enriched gluten-free pasta with microelements, especially with iron and manganese, but also with iron and zinc. The addition of banana in gluten-free pasta, both in dry and fresh form, leads to an increase in iron content, the product proved to be a functional product intended for people suffering from iron deficiency anemia. The addition of FB or DB in pasta leads to manganese fortification of products, so manganese supplementation intake in gluten-free pasta products should be considered an important outcome of the study.

The addition of banana in pasta induce an increase of TP content, the increase being significant when bananas are pretreated by adding ascorbic acid which also acts as antioxidants. The maximum amount of ferulic and coumaric acid content was recorded in the case of 20% FB addition.

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#### References

1.ABBASPOUR N., HURRELL R., KELISHADI R., Review on iron and its importance for human health, J Res Med Sci., 19 (2), 2014, p. 164 – 174.

2.ALVAREZ-JUBETE L., WIJNGAARD H., ARENDT E.K., GALLAGHER E., Polyphenol composition and in vitro antioxidant activity of amaranth, quinoa buckwheat and wheat as affected by sprouting and baking, Food Chemistry, **119**, 2010, p. 770.

3.ANSON NURIA MATEO, VAN DEN BERG ROBIN, HAVENAAR ROB, BAST AALT, HAENEN GUIDO R.M.M., Bioavailability of ferulic acid is determined by its bioaccessibility, Journal of Cereal Science, **49**, 2009, p. 296.

4.ANYASI T.A., JIDEANI A.I., MCHAU G.R., Effect of organic acid pretreatment on some physical, functional and antioxidant properties of flour obtained from three unripe banana cultivars Food Chemistry, **172**, 2015, p. 515.

- 5.ANYASI TONNA A., JIDEANI AFAM I.O., MCHAU GODWIN R.A., Effect of organic acid pretreatment on some physical, functional and antioxidant properties of flour obtained from three unripe banana cultivars, Food Chemistry, **172**, 2015, p. 515.
- 6.ARRANZ S, CALIXTO F.S., Analysis of polyphenols in cereals may be improved performing acidic hydrolysis: A study in wheat flour and wheat bran and cereals of the diet, Journal of Cereal Science, **51**, 2010, p. 313.
- 7.AURORE G., PARFAIT B., FAHRASMANE L., Bananas, raw materials for making processed food products. Trends in Food Science & Technology, **20** (2), 2009, p. 78.
- 8.BOZ H., Ferulic Acid in Cereals a Review, Czech J. Food Sci., **33** (1), 2015, p. 1.
- 9.CHONG LI CHOO, NOOR AZIAH ABDUL AZIZ, Effects of banana flour and b-glucan on the nutritional and sensory evaluation of noodles, Food Chemistry, **119**, 2010, p. 34.
- 10.DE LOURDES REIS GIADA M., Food Phenolic Compounds: Main Classes, Sources and Their Antioxidant Power, Oxidative Stress and Chronic Degenerative Diseases A Role for Antioxidants Oxidative Stress and Chronic Degenerative Diseases A Role for Antioxidants, p. 88 112.
- 11.GALLARDO C., JIMENEZ L., GARCIA CONESA M.T., Hydroxycinnamic acid composition and in vitro antioxidant activity of selected grain fractions, Food Chemistry, **99**, 2006, p. 455.
- 12.GAMBUS H., GAMBUS F., PASTUSZKA D., WRONA P., ZIOBRO R., SABAT R., MICKOWSKA B., NOWOTNA A., SIKORA M., Quality of gluten-free supplemented cakes and biscuits, International Journal of Food Sciences and Nutrition, **60** (S4), 2009, p. 31.
- 13.GROZA I., TRASCA T., RIVIS A., Mathematical model or determining the volume of abrasive susceptible of levitation, Rev. Chim. (Bucharest), **60**, no. 12, 2009.
- 14.HERNÁNDEZ-NAVA R.G., BERRIOS J., PAN J., OSORIO-DÍAZ P., BELLO-PEREZ L.A., Development and characterization of spaghetti with high resistant starch content supplemented with banana starch, Food Science and Technology International, **15** (1), 2009, p. 73.
- 15.LINTSCHINGER J., FUCHS N., MOSER H., JÄGER R., HLEBEINA T., MARKOLIN G., GÖSSLER W., Uptake of various trace elements during germination of wheat, buckwheat and quinoa. Plant Foods Hum Nutr., **50** (3), 1997, p. 223.

- 16.LIU R.H., 2007, Whole grain phytochemicals and health, Journal of Cereal Science, **46**, p. 207.
- 17.MENG F., WEI Y., YANG X.J., Iron content and bioavailability in rice, Trace Elem Med Biol., **18** (4), 2005, p. 333.
- 18.NACZKA M., SHAHIDIB F., Phenolics in cereals, fruits and vegetables: Occurrence, extraction and analysis, Journal of Pharmaceutical and Biomedical Analysis, **41**, 2006, p.1523.
- 19.NEWILAH G., NGOHBRAT P., TOMEKPE K., ALTER P., FOKOU E., ETO F.X., Effect of ripening on total polyphenol contents of musa hybrids and cultivars grown in Cameroon, Proc. IC on Banana & Plantain in Africa Eds.: T. Dubois et al. Acta Hort., **879**, ISHS 2010, p. 413.
- 20.NOOR AZIAH ABDUL AZIZ, LEE-HOON HO, BAHARIN AZAHARI, RAJEEV BHAT, LAI-HOONG CHENG, MOHAMAD NASIR MOHAMAD IBRAHIM, Chemical and functional properties of the native banana (Musa acuminate balbisiana Colla cv. Awak) pseudo-stem and pseudo-stem tender coreflours, Food Chemistry, **128** 2011), p. 748.
- 21.OVANDO-MARTINEZ M., SAYAGO-AYERDI S., AGAMA-ACEVEDO E., GO II., BELLO-PEREZ, L.A., Unripe banana flour as an ingredient to increase the undigestible carbohydrates of pasta, Food Chemistry, **113** (1), 2009, p. 121.
- 22.RODRIGUEZ-AMBRIZ S.L., ISLAS-HERNÁNDEZ J.J., AGAMA-ACEVEDO E., TOVAR, J., BELLO-PEREZ L.A., Characterization of a fiber -rich powder prepared by liquefaction of unripe banana flour, Food Chemistry, **107**, 2008, p.1515.
- 23.RYAN L., THONDRE P.S., HENRY C.J.K., Oat-based breakfast cereals are a rich source of polyphenols and high in antioxidant potential, Journal of Food Composition and Analysis, **24**, 2011, p. 929.
- 24.SARAWONG C., SCHOENLECHNER R., SEKIGUCHI K., BERGHOFER E., Effect of extrusion cooking on the physicochemical properties, resistant starch, phenolic content and antioxidant capacities of green banana flour, Food Chemistry, **143**, 2014, p. 33.
- 25.SULIBURSKA J., KREJPCIO Z., REGUŁA J., GROCHOWICZ A., Evaluation of the content and the potential bioavailability of minerals from gluten-free products, Acta Sci Pol Technol Aliment., **12** (1), 2013 p. 75.
- 26.VERMA B., HUCL P., CHIBBAR R.N., Phenolic acid composition and antioxidant capacity of acid and alkali hydrolysed wheat bran fractions, Food Chemistry, **116**, 2009, p. 947

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